

BNN27 NANOFORMULATIONS FOR NOSE-TO-BRAIN DELIVERY

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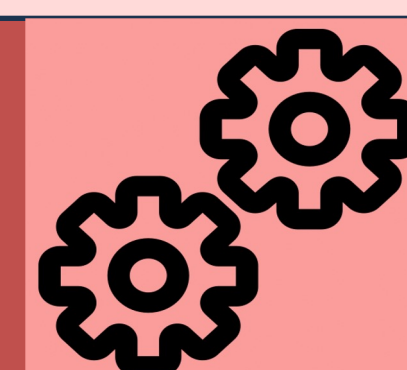
INTRODUCTION



The treatment of neurodegenerative diseases is still an unmet medical need. Small, lipophilic molecules with strong neuroprotective properties, such as BNN27 (DHEA derivative), represent new non-toxic, synthetic compounds for therapeutic applications in brain diseases [1]. Their low solubility and targeted transport to the brain are challenges for optimization of their clinical application.

Intranasal delivery offers an alternative and promising way for brain targeting. In this project, we studied the design of two innovative mucoadhesive nanoformulations for intranasal delivery of BNN27: i) negatively-charged liposomes (LIP) coated with chitosan (CHT) and ii) mucoadhesive nanoemulsions (BNE) with CHT.

METHODS



BNN27-loaded LIP with lipid composition PC or PC/PG 9:1 mol/mol were prepared with thin film hydration method. LIP were coated with CHT by dropwise addition of CHT solution in the liposome dispersion. BNE were prepared by the spontaneous emulsification method using Capmul MCM as the oily phase, Tween 80 as surfactant and Transcutol:Propylene Glycol (1:1) as co-surfactants, according to solubility studies [2]. Size distribution and ζ -potential were measured by dynamic light scattering (DLS) and laser Doppler electrophoresis (LDE), respectively.

For the monolayer studies, hCMEC/D3 cells were seeded on Transwell filters, pre-coated with type I collagen [3] and Rhodamine (Rho)-labeled nanoformulations were used.

For the *in vivo* study in mice, 25 μ L/mouse of each BNN27-formulation was administered into each nostril. After sacrifice, brains were quickly collected and frozen. BNN27 quantification were performed with HPLC-MS [2].

RESULTS



After preformulation studies [2], medium MW CHT-coated PC/PG/BNN27 liposomes at 0.1 w/w CHT/lipid ratio demonstrated highest coating efficiency (table 1) and mucoadhesive properties (Figure 1a). Furthermore, BNEs with 8% w/w or 10% w/w Capmul MCM and 0.3% w/w CHT, displayed optimal physicochemical characteristics, compared to BNEs with Carbopol (CAR) as mucoadhesive agent (Figure 1b). The optimal BNN27 nanoformulations were evaluated for their permeability across a cellular BBB model (hCMEC/D3 cell monolayers) *in vitro* (Figure 2). The translocation of NE-associated Rho and the corresponding permeability values were significantly higher than the corresponding values of LIPs. The *in vivo* fate of the innovative nanoformulations after intranasal administration in mice revealed the superiority of BNE with CHT for more efficient distribution of BNN27 to the brain, compared to liposomal BNN27 (Figure 3).

Table 1: Physicochemical properties of the CHT-coated LIPs. Effect of lipid membrane composition and CHT type and amount.

Lipos	CHT MW-CHT/LIP (w/w)	Mean Diameter (nm)	PDI	ζ -Potential (mV)	Coating Efficiency (%)
PC	Non-coated	78.1 \pm 1.15	0.281	-3.48 \pm 0.49	-
	Low-0.1	123.7 \pm 2.17	0.398	3.65 \pm 0.27	3.5 \pm 0.2
	Med.-0.1	158.8 \pm 6.27	0.405	4.57 \pm 0.18	4.8 \pm 0.6
PC/PG	Non-coated	83.01 \pm 1.36	0.263	-12.4 \pm 0.689	-
	Low-0.1	822 \pm 21.54	0.452	15.2 \pm 0.47	79.9 \pm 1.5
	Med.-0.1	1147 \pm 86.27	0.447	23.1 \pm 1.41	85.1 \pm 4.5
	Med.-0.3	2563.5 \pm 109.6	0.592	24.4 \pm 1.273	78.2 \pm 0.7
	Med.-0.5	5155 \pm 51.24	0.617	29.4 \pm 0.689	81.4 \pm 1.8

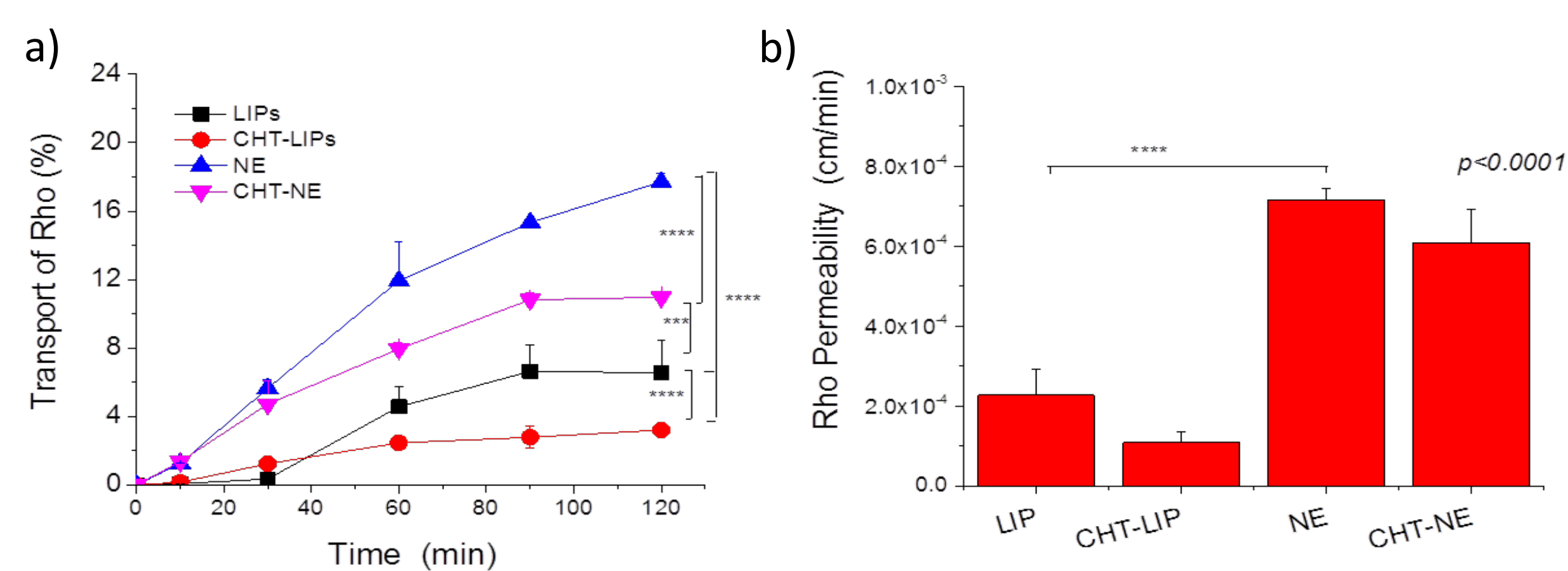


Figure 2: a) Transport of formulation-associated Rho (%) across the monolayer with the time (following incubation with LIP and NE formulations at 0.2 μ M Rho concentration). b) Permeability values of formulations (formulation-associated RHO permeability) across the hCMEC/D3 monolayer (calculated from the results of Figure 1a).

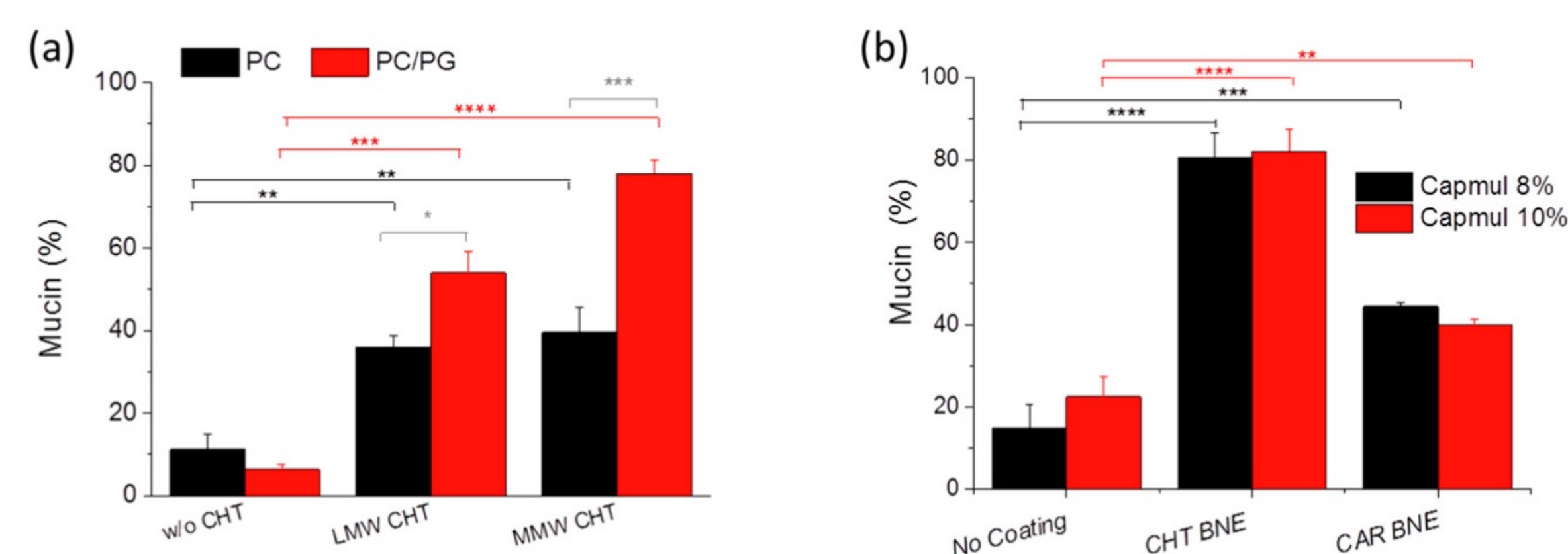


Figure 2: Mucoadhesive properties (expressed as the capability to adsorb mucin (% adsorbed) of: (a) CHT-coated LIPs. (b) CHT- or CAR-coated NEs. Non-coated formulations were evaluated under identical conditions (in all cases) for comparison.

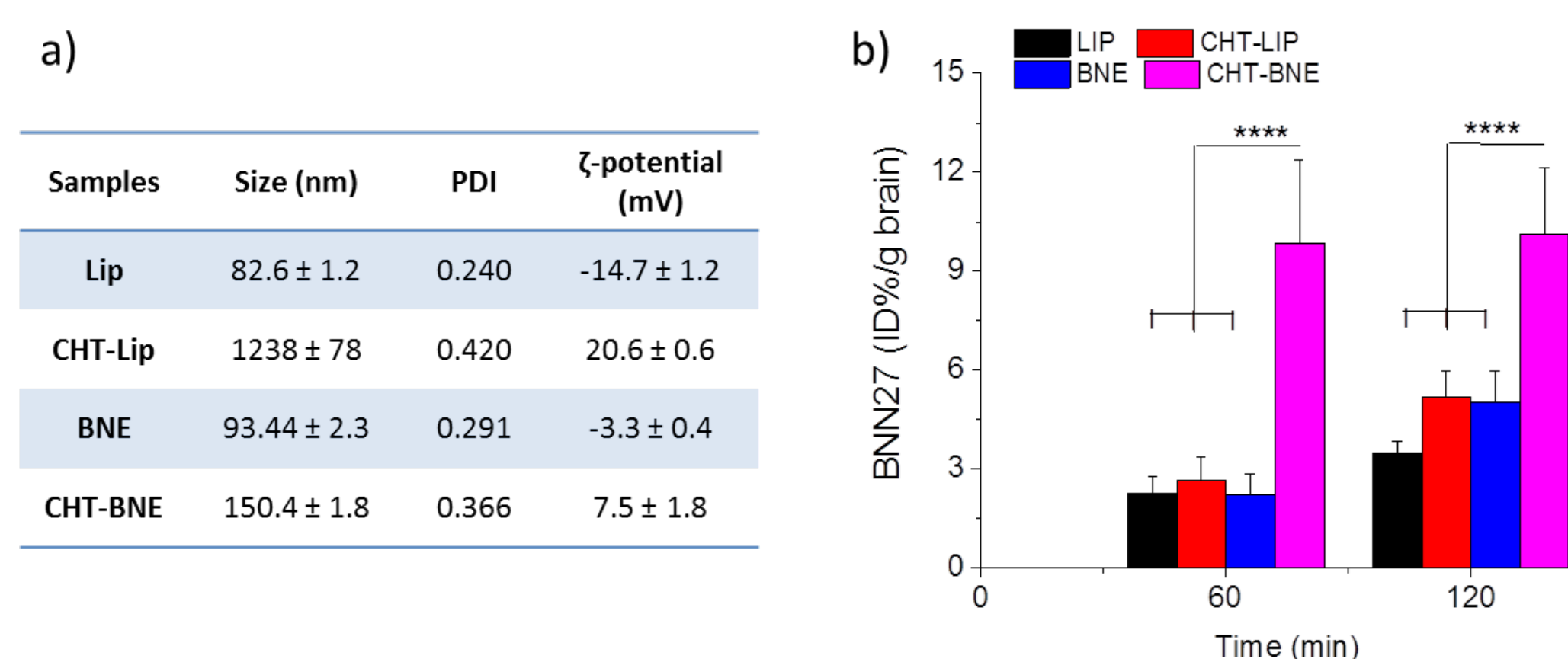


Figure 3: a) Physicochemical properties of the formulations used in the *in vivo* disposition study. b) Brain disposition of BNN27 1 and 2 h post-intranasal administration of the BNN27 loaded liposomal and NE formulations.

CONCLUSIONS



CHT-BNE formulation developed herein was demonstrated to confer faster and higher nose-to-brain delivery of BNN27 compared to CHT-LIPs. Such NEs could be considered as alternative systems for the brain delivery of lipophilic drugs following intranasal administration. Nevertheless, extended biocompatibility and toxicity studies are required to exclude any potential toxicity issues in relation to the cytotoxicity differences between the LIPs and NEs above-mentioned due to the high surfactant content of NEs.

REFERENCES

- [1] A. Gravanis et al. *Oncotarget*, 2017, 8, 9005-9006
- [2] M. Kannavou et al. *Pharmaceutics*, 2023, 15, 419
- [3] E. Markoutsas et al. *Pharmaceutical Research*, 2014, 31, 1275-1289

Acknowledgments

This research was co-financed by the European Regional Development Fund of the European Union and Greek national funds through the Operational Program Competitiveness, Entrepreneurship and Innovation, under the call RESEARCH-CREATE-INNOVATE (project code: T1EDK-03186, Acronym: DINNESMIN).

